LUNG CYTOTOXICITY OF COMBINED EXPOSURE TO REFRACTORY CERAMIC FIBRES AND CIGARETTE SMOKE

Silvia Černá, Marta Hurbánková, Zuzana Kováčiková, Milan Beňo, Soňa Wimmerová

Slovak Medical University, Bratislava, Slovak Republic
e-mail: silvia.cerna@szu.sk

Received: June 10, 2005; Accepted: September 25, 2005

Key words: Lungs/Bronchoalveolar lavage/Refractory ceramic fibres/Smoking/Cytotoxicity/ Lysosomal enzymes

Changes in some lung cytotoxic parameters after exposure to refractory ceramic fibres (RCF) or to cigarette smoke (S) and after combined exposure to RCF+S were studied in male Wistar rats in order to evaluate their potential adverse health effects. Four groups of rats were treated as follows: 1) intratracheally instilled by saline solution (0.4 ml); 2) intratracheally instilled by 4 mg of RCF; 3) exposed only to S (85 mg of total particulate matter/m³ air) for two hours daily; 4) exposed to RCF+S. After 6 months the animals were exsanguinated and the bronchoalveolar lavage (BAL) was performed. Viability and phagocytic activity of alveolar macrophages (AM), activity of lactate dehydrogenase (LDH) in cell-free BAL fluid (cfBALF), acid phosphatase (ACP) and cathepsin D (CATD) in cfBALF, in BALF cells and in the lung tissue were estimated. Viability of AM was depressed by every type of exposure with RCF+S effect being at least additive. Phagocytic activity of AM increased in the presence of RCF. No significant changes in LDH activity were found. Activities of lysosomal enzymes measured in the lung tissue homogenates were not significantly changed, but those in the cfBALF increased especially after exposure to S with most expressive increase in BALF cells after exposure to S and RCF+S. In the case of CATD the effect of RCF+S was more than additive. The results point out to the persistence of the RCF exposure cytotoxic effects and their amplification by cigarette smoke.

INTRODUCTION

RCF – amorphous or partially crystalline materials made from kaolin clay or oxides of aluminium or other metal oxides have many properties (e.g. low heat storage, low thermal conductivity, resistance to thermal shock, chemical resistance) supporting their utility as an excellent isolating material. Nowadays, after banning of asbestos use in most of the countries, RCF together with other man made vitreous fibres (MMVF) are getting into centre of interest. They are used as asbestos substitutes excepting their lower impact on the human and environmental health. Currently produced RCF contain fibres with the diameter distribution within the respirable range. In vitro studies of RCF dissolution showed, that RCF are dissolved more rapidly than amosite fibres, but slower than other MMVF. RCF belong to fibres with long biodurability comparable with amosite and dependent on the length of fibres. It is supposed that the clearance of short fibres is accomplished primarily by macrophages and that of long fibres by dissolution and disintegration. Regarding to the mentioned properties (respirability, biodurability) a negative influence of RCF exposure on the respiratory system can be expected. Studies on rodents including the inhalation studies showed that exposure to higher doses of RCF resulted in pulmonary fibrosis and increases of the number of lung tumours and mesothelioma. RCF are generally considered to be rodent carcinogens. International Agency for Research on Cancer inserted RCF into Group 2B of carcinogenic risks to humans – possibly carcinogenic. According to the results of epidemiological studies summarised in the work of Venturin et al. exposure to RCF caused elevated incidence of pleural plaques without evidence of fibrosis, mesothelioma or incremental lung tumours. Cowie et al. found some association between small opacities and cumulative exposure to RCF. Inverse relation between pulmonary function and exposure to fibres was found only in smokers. A negative synergism between RCF exposure and smoking might be expected. The aim of our work was to follow the cytotoxic effect on lung tissue after experimental exposure of rats to combined RCF and cigarette smoke.

MATERIAL AND METHODS

The used RCF fibres (Carbolane fibres – Saint Gobain Ceramiques Industrielles, Nanterre, France) including their characterisation were kindly supplied by Dr. É. Tatrai, Fodor Jozsef National Center for Public Health, Budapest, Hungary. The length and diameters of them are in Table 1.

Male Wistar rats (VELAZ, Prague, Czech Republic) weighing at the beginning of the experiment 152.9 ± 6.6 g were divided into 4 groups and treated as follows: 1. group
(control) – intratraehally instilled by 0.4 ml of saline solution administered in two 0.2 ml doses; 2. group – intratracheally instilled by 4 mg of refractory ceramic fibres (RCF) administered in two 2 mg doses in the form of suspension in saline solution (1 mg RCF/0.1 ml saline solution); 3. group - exposed to diluted mainstream cigarette smoke (S) at the concentration of 85 mg of total particulate matter (TPM)/m³ air for two hours daily (except Saturdays and Sundays) in a whole-body exposure chamber (THRI, Lexington, USA) reaching this concentration by burning 8 standard research cigarettes 1R1 type (THRI, Lexington, USA); 4. group – intratracheally instilled by 4 mg RCF (as group 2) and exposed to S (as group 3). Six months after beginning of the exposure the animals were anaesthesized by thiopental (150 mg/kg) and exsanguinated by cutting the vena cava caudalis. Bronchoalveolar lavage (BAL) was performed by modified method of Myrvik. After estimation of the viability of AM the bronchoalveolar lavage fluid (BALF) was centrifuged at 400 g for 10 minutes at 4 °C, the cell-free BALF (cfBALF) was transferred into clean cooled glass tubes and the cell sediment adjusted by sterile saline solution to 1×10⁶ cells/ml. The lungs were perfused and 10% stock homogenate in phosphate buffered saline (PBS) was prepared. Lactate dehydrogenase (LDH) was measured immediately (LD 105 UV Lachema Brno, Czech Republic). Acid phosphatase (ACP) in cfBALF (Acid phosphatase AC 565, Randox Laboratories, Antrim, UK) was measured during the day of BAL performance. The rest of cfBALF, the cell sediments and the lung tissue homogenates were stored at –70 °C until they were analysed. The cell sediments were resuspended in TRITON X100 dissolved in PBS and the tissue homogenates were rehomogenized in Triton X100 (final concentration of TRITON in a sample was 0.1 %)³. After 3 times repeated freezing and thawing the treated cell sediments and tissue homogenates were centrifuged at 15 000 g for 20 minutes at 4 °C. Activities of the lysosomal enzymes were measured in the supernatants.

Mann-Whitney test was used for the comparison of values from chosen groups.

RESULTS AND DISCUSSION

Viability of AM significantly decreased after all types of exposure (Fig. 1). This indicated the duration of cytotoxic effect of RCF and confirmed the expected cytotoxicity of cigarette smoke. Exposure to cigarette smoke amplified the negative effect of RCF more than additively. Exposure to S alone, similarly as in our previous work.

Table 1. Distribution of ceramic refractory fibres.

<table>
<thead>
<tr>
<th>fibre diameter (µm)</th>
<th>%</th>
<th>fibre length (µm)</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 1</td>
<td>6</td>
<td>20–40</td>
<td>17</td>
</tr>
<tr>
<td>= 1</td>
<td>12</td>
<td>41–60</td>
<td>11</td>
</tr>
<tr>
<td>&lt; 3</td>
<td>35</td>
<td>61–80</td>
<td>14</td>
</tr>
<tr>
<td>= 3</td>
<td>8</td>
<td>81–100</td>
<td>14</td>
</tr>
<tr>
<td>&gt; 3</td>
<td>39</td>
<td>&gt; 100</td>
<td>44</td>
</tr>
</tbody>
</table>

Composition of fibres: SiO₂ 45-60%; Al₂O₃ 40–55%

Table 2. Activity of lactate dehydrogenase, acid phosphatase and cathepsin D in cell-free bronchoalveolar lavage fluid and lung tissue homogenates

<table>
<thead>
<tr>
<th>enzyme</th>
<th>C</th>
<th>RCF</th>
<th>S</th>
<th>RCF + S</th>
</tr>
</thead>
<tbody>
<tr>
<td>LDH jkat.g protein⁻¹</td>
<td>6.47 (4.22; 7.82)</td>
<td>5.31 (3.96; 6.99)</td>
<td>7.56 (7.12; 8.45)</td>
<td>6.01 (5.86; 6.48)</td>
</tr>
<tr>
<td>ACP nkat.g protein⁻¹</td>
<td>59.97 (54.29; 69.07)</td>
<td>61.13 (49.27; 83.64)</td>
<td>90.65 * ↑ (83.40; 96.37)</td>
<td>67.39 (58.63; 78.82)</td>
</tr>
<tr>
<td>CATD Utyr. µg protein⁻¹</td>
<td>63.39 (57.54; 67.66)</td>
<td>67.95 (62.12; 77.29)</td>
<td>95.69* ↑ (83.95; 124.09)</td>
<td>94.06* ↑ (91.71; 105.15)</td>
</tr>
</tbody>
</table>

Values represent medians and 25th and 75th percentiles.

C – control group, RCF – group exposed to refractory ceramic fibres, S – group exposed to cigarette smoke, RCF + S – group exposed to refractory ceramic fibres and cigarette smoke, LDH – lactate dehydrogenase, ACP – acid phosphatase, CATD – cathepsin D, Utyr. µg of tyrosine released in an hour time. Comparison with control group: * P < 0.05 , ↑ – increase.
significantly depressed the phagocytic activity of AM (Fig. 2). Contrary to amosite, the tested RCF stimulated the phagocytic activity.

Enzyme activities in cBALF and in lung tissue homogenates are shown in Table 2. Increase in the activity of LDH – a cytosolic enzyme – and in the activity of lysosomal enzymes in extracellular fluids are considered to be the consequence of the leakage of these enzymes from the damaged tissue and in the case of cBALF also of the leakage from AM. Activity of LDH in this experiment was not significantly influenced by any of tested exposures (Table 2). We measured the LDH activity six months after instillation. Results of the report of inhalation study with RCF in rats showed that the differences in LDH activities between control and exposed groups disappeared between the 31st and 93th day after end of RCF inhalation. Significantly higher activities of lysosomal enzymes in cBALF were found only after exposure to S alone or in the case of CATD also after combined exposure (Table 2).

Exposure to RCF alone did not change the ACP activity in BALF cells (Fig. 3). This fact together with the similarity of significant increase in ACP activity after exposure to S and to RCF+S indicates that the influence of smoke on the ACP activity was much more expressive than the influence of RCF. Significant increase of cathepsin D activity after all types of exposure confirmed the sensitivity of this parameter at the evaluation of lung cytotoxicity of examined respirable substances. In combined exposure the effect of examined substances was more than additive. Most of the articles dealing with the activities of lysosomal enzymes in AM after in vivo or in vitro exposure to respirable substances describe the intracellular and extracellular concentration or activity of these enzymes after incubation of the isolated AM in the medium. Increased (often dose dependent) leakage into the medium (and consequently decreased activities in the intracellular space) are mentioned. We did not incubate the isolated BAL cells because we did not want to monitor the leakage of lysosomal enzymes. The activities of lysosomal enzymes we measured were the activities at the time of BAL performance. The most expressive changes were found in the cathepsin D activity (Fig. 4). This aspartyl endopeptidase is closely associated with tumour progression in some human malignancies including
lungs, and it is overexpressed in inflammatory status and modulated by inflammatory mediators. Cigarette smoke was described as a potent inducer of this enzyme activity in AM.

Examination of the effect of RCF cigarette smoke on cells in lung tissue results into the following conclusions: (i) The expected cytotoxicity of RCF was confirmed by the significant decrease in viability of AM, significant increase in CATD activity in cfBALF, and significant increase in ACP and CATD activity in BALF cells; (ii) Cytotoxic activity of cigarette smoke was demonstrated by the significant decrease in viability of AM, significant decrease in phagocytic activity of AM, significant increase of ACP and CATD activity in cBALF, and significant increase of ACP and CATD activity in BALF cells. (iii) Exposure to cigarette smoke amplified the negative effect of RCF. (iv) Cathepsin D activity was the most sensitive indicator of cytotoxic effects of tested substances.

ACKNOWLEDGEMENT

The authors wish to express their thanks to Mrs Helena Bobeková, Mrs Mária Valentová, Ing. Marta Staruchová, RNDr. Katarina Volková PhD, Mrs. Darina Cepcová, Mrs. Jamila Jantsková and Miss Lubica Miklošková for their excellent assistance.

REFERENCES