Glucose concentrations in blood and tissue – a pilot study on variable time lag

Rudolf Chlupa a,b,c, Jan Krejci d, Mark O´Connell e, Blanka Sebestova a, Robert Plicka a, Lucie Jezova a, Tereza Brozova a,d, Blanka Doubravova a,d, Hana Zalesakova a,d, Emilia Durajkova a,d, Jiri Vojtek d, Josef Bartek a

Aim. The aim of this pilot study was to acquire insight into the parameters of glycaemic control, especially, (1) the time delay (lag phase) between plasma and tissue glucose concentrations in relation to rise and fall in glucose levels and (2) the rate of glucose increase and decrease.

Methods. Four healthy people (HP), 4 people with type 1 diabetes (DM1) and 4 with type 2 diabetes (DM2) underwent concurrent glucose measurements by means of (1) the continuous glucose monitoring system (CGMS-Medtronic, Medtronic-Minimed, CA, USA, calibrated by the glucometer Calla, Wellion, Austria, and, (2) the Beckman II analyser to measure glucose concentrations in venous plasma. Samples were taken on 4 consecutive days in the fasting state and 4 times after consumption of 50 g glucose. Carelink Personal, MS Excel, Maple and Mat lab were applied to plot the evolution of glucose concentration and analyse the results. The time difference between increase and decrease was calculated for HP, DM 1 and DM 2.

Results. In DM1 and DM2, glucose tolerance testing (GTT) resulted in slower transport of glucose into subcutaneous tissue than in HP where the lag phase lasted up to 12 min. The maximum increase/decrease rates in DM1 and DM2 vs HP were 0.25 vs <0.1 mmol/L/min.

Conclusion. CGMS is shown to provide reliable plasma glucose concentrations provided the system is calibrated during a steady state. The analysis of glucose change rates improves understanding of metabolic processes better than standard GTT.

Key words: diabetes mellitus, continuous glucose monitoring, lag phase, glucose transport, calibration

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INTRODUCTION

Euglycaemic control is of paramount importance in diabetes especially in relation to preventing late diabetes complications. Several markers of glycaemic control have been used in routine practice as well as in clinical trials. Apart from clinical investigation, laboratory parameters such as quantitative glycosuria, ketonuria, fasting glycaemia, glycated serum proteins, 1,5 anhydroglucitol, and concentration of haemoglobin A1c in blood are used to assess carbohydrate metabolism.

Above all plasma glucose concentrations over 24 h recorded as mean plasma glucose and glycaemic variability over 24 - 72 h (ref.3) have become the principle parameters for guiding treatment and assess the efficacy of drugs on patient glycaemic control.

From 1974, following the introduction of glucometers into daily routine, self-monitoring of plasma/ blood glucose (SMPG/SMBG) became the predominant mode of glucose monitoring. From the year 2000, the use of continuous glucose monitoring systems (CGMS) has shown remarkable benefits for both people with type 1 and type 2 diabetes. The development of CGMS technology is a great step toward modern diabetes management. CGMS has also been used to investigate the glycaemic indexes of food at different times of the day. This overcomes the limitations of traditional monitoring by producing glucose profiles instead of distinct measurements, real-time glucose values, glucose trends and warnings when glucose values approach dangerously low or high concentrations. In this way, frequent use of CGM systems and careful pattern analysis can improve glycaemic control.

On the other hand, evaluation of the prolific data provided by CGMS often makes the implementation of CGMS in practice, difficult. The relationship between plasma glucose concentration and interstitial glucose dynamics had to be established. Rebrin concluded that differences between plasma and interstitial glucose will not be a significant obstacle in advancing the use of interstitial fluid as an alternative to plasma glucose measurements. Thennadil compared glucose concentration in interstitial fluid and capillary and venous blood during rapid changes...
in blood glucose concentrations\textsuperscript{17}. Boyne found the time lag of interstitial glucose behind plasma glucose, regardless whether glycaemia was rising or falling\textsuperscript{18}.

CGMS calibration is influenced by the speed (time rate [mmol/L/min]) of glucose transport between plasma, interstitium and intracellular space. Current CGMS methods derive glucose concentrations from different tissues using subcutaneous/"interstitial" values of electrical current (ISIG – Input Signal for Glucose) for measurement and whole blood/plasma for calibration. Correct calibration of subcutaneous sensors is mandatory for reliable CGMS results. However, correct calibration can be obtained only if the concentrations in plasma and tissue are the same. The measured values can be affected by several factors such as accuracy and precision of the calibrating system, variability of plasma glucose concentration resulting from food consumption or stress and muscular exercise.

The aim of this pilot study was to gain insight into the parameters of glycaemic control, especially, (1) the time delay (lag phase) between postprandial glucose concentrations in venous plasma (measured by the analyser Beckman II) and subcutaneous tissue (measured by CGMS - Medtronic calibrated by the Calla glucometer-stripe system) as they increase and decrease in healthy people (HP), in persons with diabetes type 1 (DM1) and in people with diabetes type 2 (DM2), and, (2) the rates of glucose concentration increase and decrease.

MATERIALS AND METHODS

Tested volunteers

Twenty five volunteers for this pilot study (HP, DM1 and DM2) were recruited from students, hospital staff and patients at diabetes centres. Informed consent according to the most recent Helsinki Declaration was signed by all participants. The purpose and procedures of the research protocol were explained. Their rights to refrain from the trial under any circumstances and for any reason were emphasized. Clinical and laboratory investigations of all volunteers were performed on admission to day-4 of the stay in the diabetes department to confirm that they all met the criteria required for the trial (age above 18 years old, no active infection no acute disorder within 3 weeks before the trial, no haemorrhagic disorder, no malignancy, no pregnancy, no oral or injectable steroids).

All the participants agreed to the following prerequisites:

- No smoking or alcohol over a period of one day before and during the 4 days of study.
- Willing to keep to the defined protocol over the whole study period.
- Carrying on with usual medication and insulin administration (if necessary).

People with frequent disturbances of function of venous catheter and serious deviations from the study protocol were excluded such that only twelve volunteers with complete data sets were considered for final evaluation: HP (n = 4, age 21 - 46 y, BMI 22.0 - 34.7 kg/m\textsuperscript{2}, HbA1c 34 - 35 mmol/mol); DM1 (n = 4, age 37 - 66 y, duration of DM1 0 - 42 y, insulin 23 - 42 IU/d, BMI 21.1 - 26.0 kg/m\textsuperscript{2}, HbA1c 60 - 81 mmol/mol); DM2 (n = 4, age 57 - 68 y, duration of DM2 up to 23 y, insulin up to 60 IU/d, BMI 21.7 - 38.0 kg/m\textsuperscript{2}, HbA1c 50 - 88 mmol/mol).

Equipment and software

Standard devices and programmes were:

(1) Continuous glucose monitoring system (CGMS), namely, subcutaneous sensor Enlite, for measuring the ISIG transferred by means of an external transmitter Minilink from interstitium to the monitor Guardian where it was automatically converted (using calibrating algorithm) to the values of plasma glucose concentrations, Medtronic Minimed, Northridge, CA, USA.

(2) Glucometer system Calla Premium and glucose oxidase strips (Wellion, Austria) for measuring values of glucose concentrations in capillary plasma to calibrate the CGMS.

(3) Beckman analyser typ II to estimate glucose concentrations in venous plasma.

Software programs Carelink Personal v. 3.0 (Medtronic Minimed, Northridge, CA, USA), MS Excel, Maple and Mat lab were applied to plot the evolution of glucose concentration obtained by CGMS and by Beckman analyser during the whole study, respectively.

Study design

This was a cross-sectional trial performed in the Diabetes Department of the Institute of Neurology and Geriatrics, Moravský Beroun, Czech Republic from October 2013 to April 2014. Approval of the Ethics Committee of the University Hospital Olomouc and Faculty of Medicine and Dentistry, Palacky University Olomouc was obtained.

Investigation of each volunteer lasted 4 days (from Friday to Monday). On the first day, one or two volunteers were admitted to one ward and remained there under the scheduled check-ups of investigators and continuous supervision of one specialized nurse until the fourth day.

On admission (Day 1), a thorough clinical investigation was performed.

Sensor Enlite was inserted by a specialized nurse into the subcutaneous tissue of the abdomen or arm and within 15 min was connected to a transmitter Minilink. The first CGMS calibration was performed two hours later (at about 10:00 h, still in the fasting state) using the value of capillary plasma glucose concentration from glucometer Calla which was manually put into the monitor Guardian. Next, the CGMS was operated according to the instructions in the Medtronic Manual. All values of glucose concentration from Calla (usually 18 per day) were put into Guardian.

On Day 1, 2, 3 and 4 in addition to the CGMS, blood samples from cubital vein were obtained using an indwelling catheter which was inserted on the first day.
and remained functioning over the whole study period or changed only if any malfunction or discomfort occurred. Venous blood was taken in the fasting state at 6:00 h and then each hour till 24:00, and on day 4 from 6:00 till 11:00. This venous blood was centrifuged within 5 min after the drawing at 2000 rev/min, for 5 min. Plasma was immediately analysed or (if taken between 14:00 and 24:00 h) stored at -18 °C for analysis the next day. Glucose concentrations in venous plasma were measured in all sample tubes using the Beckman analyser. Fifty grams of glucose dissolved in 300 mL tea or water was administered in the fasting state as a breakfast usually at 7:00 h. Following this 2-h oral glucose tolerance test (OGTT), all volunteers received standard portions of 4 different meals served at 4-hourly intervals (i.e. at 11:00 h, 15:00 h, 19:00 h and 23:00 h). Each portion contained 50 g of absorbable carbohydrates and adequate amount of proteins: 260 g carbohydrates, 70 g proteins and 88 g fat (i.e. a total of 8930 kJ) were consumed daily on day 1, 2 and 3.

On day 4 the study ended after the OGTT at 11:00 h. The meal plan is defined in Table 1. No additional meals or drinks were allowed.

Data processing and statistic analysis
The data from the monitor Guardian were downloaded to the therapy management software for diabetes Carelink Personal (www.carelink.minimed.com) and then exported to MS Excel to plot the daily evolution of glucose concentration obtained by CGMS and by Beckman analyser, respectively. Data from the Beckmann analyser were included manually.

For each volunteer, the time difference between increase and decrease was estimated and averaged for each HP, for DM1 and for DM2, respectively.

The data analysis was performed by our own programs created in Maple, Matlab and MS Excel. The programs were created using standard Maple and Matlab libraries and linked to Excel (graphical output). Data were processed using splines of the 3rd order.

<table>
<thead>
<tr>
<th>Day</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day</td>
<td>Run-in day</td>
<td>Study day</td>
<td>Study day</td>
<td>Study day</td>
</tr>
<tr>
<td>Breakfast 7:00</td>
<td>Training – GTT</td>
<td>GTT 1</td>
<td>GTT 2</td>
<td>GTT 3</td>
</tr>
<tr>
<td>Lunch 11:00</td>
<td>Glucose (50 g)</td>
<td>Glucose (50 g)</td>
<td>Glucose (50 g)</td>
<td>Glucose (50 g)</td>
</tr>
<tr>
<td>Snack 15:00</td>
<td>Multigrain Rye</td>
<td>Rolled Wafers</td>
<td>Multigrain Rye</td>
<td>End of the study</td>
</tr>
<tr>
<td>Dinner 1 19:00</td>
<td>Knäckebröt</td>
<td>with vanilla-cinnamon</td>
<td>Knäckebröt</td>
<td>at 11.00 h</td>
</tr>
<tr>
<td>Dinner 2 23:00</td>
<td>Yogurt Florian</td>
<td>Yogurt Florian</td>
<td>Ham + Multigrain Rye</td>
<td>Rolled Wafers</td>
</tr>
<tr>
<td></td>
<td>+ Multigrain Rye</td>
<td>Multigrain Rye</td>
<td>Knäckebröt</td>
<td>with vanilla-cinnamon</td>
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<td></td>
<td>Knäckebröt</td>
<td>Knäckebröt</td>
<td>Knäckebröt</td>
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</tr>
</tbody>
</table>

The results were presented using analysis of first derivative of glucose profile as the first approximation step of process description.

Estimation of time delay between plasma and tissue and glucose increase/decrease rates
The time delay between plasma and tissue during plasma glucose concentration increase (BT↑) was measured as time between maximum glucose concentration increase rates in plasma and tissue. Maximum glucose concentration increase in tissue was measured by CGMS. Similarly, the time delay at glucose concentration decrease (TB↓) was measured as the difference in CGMS and Beckman values at maximum decrease rates in tissue (measured by CGMS) and plasma glucose decrease rate. Both quantities are well defined from the mathematical point of view. BT↑ is defined as the time difference between inflex points of plasma and tissue glucose concentration when the glucose concentration increases. TB↓ is defined in the same manner when the glucose concentration in venous plasma decreases.

Capillary plasma glucose concentration was measured by the glucometer-strip (glucoseoxidase) system Calla and on the analyser Beckman II using glucoseoxidase enzyme. The best correlation between Beckman analyser and system Calla proved the accuracy of plasma glucose measurement and correct CGMS calibration using glucometer Calla.

RESULTS
Time delays between tissue and plasma are displayed in Table 2.

The result for the reliable function of CGMS is shown in Fig. 1 as an analysis of glucose increase/decrease trends. An excellent agreement in the trends of CGMS and blood/plasma is seen. The relation between trend in tissue and blood/plasma is approximately same and the same as responses to glucose load.
Table 2. Delay between glucose concentration in tissue and plasma during the increase of plasma glucose (BT↑) and decrease of plasma glucose (TB↓) in HP, DM1 and DM2.

<table>
<thead>
<tr>
<th>Person</th>
<th>BT↑ [min]</th>
<th>TB↓ [min]</th>
</tr>
</thead>
<tbody>
<tr>
<td>No</td>
<td>Mean</td>
<td>SD</td>
</tr>
<tr>
<td>HP</td>
<td>P08</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>P21</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>P22</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>P24</td>
<td>12</td>
</tr>
<tr>
<td>DM 1</td>
<td>P06</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>P11</td>
<td>8</td>
</tr>
<tr>
<td></td>
<td>P17</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>P18</td>
<td>18</td>
</tr>
<tr>
<td>DM 2</td>
<td>P01</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>P02</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>P15</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>P23</td>
<td>7</td>
</tr>
</tbody>
</table>

(- | -) no agreement between trends in CGMS (tissue ISIC calibrated for capillary plasma) and Beckman (venous plasma)
(x | -) one agreement between trends in CGMS (tissue ISIC calibrated for capillary plasma) and Beckman (venous plasma)

The analysis of glucose increase/decrease (GID) rates is demonstrated in Fig. 2 - 4.

Fig. 1. Comparison of the Beckman and CGSM-Medtronic trend results.

Fig. 2. Development of positive and negative trends in glycaemia of a healthy woman (P21).

Fig. 3. Development of positive and negative trends in glycaemia of a man with DM2 (P01).

Fig. 4. Development of positive and negative trends in glycaemia of a man with newly diagnosed DM1 (P18) on intensive insulin treatment.

The analysis of glucose increase/decrease (GID) rates is demonstrated in Fig. 2 - 4.

Fig. 2 shows the trends of GID in a healthy woman. The glucose controlling mechanism is able to compensate the glucose changes in an effective manner which results in very small values of GID. The average glucose concentration during the test was 6.4 mmol/L (SD 1.25 mmol/L). The 3rd GTT was fully compensated by the body glucose control mechanisms. No GID higher than 0.06 mmol/L/min was detected at the third GTT. No change
rates could be recognized at the fourth GTT, which may be the result of excellent function of the B-cells.

Fig. 3 shows the trends of GID in a man with DM2. The average glucose concentration is 8.6 (SD 1.98) mmol/L. Repetition of GTT starts to disrupt glucose metabolism (day 2) and the worsening continues on days 3 and 4. This phenomenon was observed in 10 volunteers (healthy, DM1, DM2) out of 12.

Fig. 4 shows the GID of a volunteer with recent DM1 on insulin. At the beginning (day 1), his control is quite good, and the pancreas seems to function adequately. The GID response in the course of GTT was about 0.1 mmol/L/min which is comparable to a healthy person. However, as early as on day 2, the GTT load could not be adequately controlled either by injections of exogenous insulin or by the resting function of the pancreas. The glucose increases at a rate of 0.24 - 0.26 mmol/L/min and this increase is followed by a large decrease ranging from 0.14 mmol/L/min after the first GTT to 0.25 mmol/L/min after the third GTT.

DISCUSSION

This pilot study provides insight into important parameters of glycaemic control. The outcomes support the value of precise estimation of time lag and change rates between glucose concentrations in plasma and interstitium in the course of rising and falling. In this way it contributes to improving diagnostic procedures and the efficacy of diabetes treatment.

Although the results are limited due to the small sample size they provide valuable data for improving the standard of diabetes therapy and the design of coming studies.

The threshold (limit) of GID rates between healthy and DM1 and DM2 volunteers was found to be 0.1 mmol/L/min. This value is the result of the present pilot study but needs to be confirmed by studies on larger numbers of volunteers.

The importance of CGMS in effective therapeutic outcomes has been clearly demonstrated by a number of previous studies. On the other hand, in some cases low reliability of CGMS in measuring the glucose concentration change rates is also reported. These contradictory issues cannot be neglected. In people with diabetes, the glucose control system of the body needs hours to recover after glucose intake. A stimulus can initiate the oscillations. A stimulus can initiate the oscillations.

The CGMS in this study (using the monitor Guardian) was applied to monitor the state of the patient, even though recent versions of monitors and insulin pumps include processing of trends of increase/decrease of glucose concentrations which distinguish change rates of 0.11 mmol/L/min (marked on the display by one arrow) and of 0.22 mmol/L/min (marked by two arrows).

If all experimental conditions comply with ideal measuring conditions, excellent agreement between CGMS and Beckman is found (Fig. 1). However, such agreement was found only in 1/12 volunteers. Table 2 shows quite convincingly that the lag time between blood glucose and interstitial fluid glucose varies considerably between cases. Possible reasons for the poor correlation between these two compartments include physiological factors such as temperature, blood perfusion, osmolality, actual glucose concentrations in intracellular vs. extracellular space, activity of glucose transporters, insulin concentration, state of the insulin receptors etc. These factors may vary between individuals and also be different at specific sites of the body. The potential influence of medication must also be considered.

This discussion confirms the importance of the development of a reliable and robust system which can measure the glucose concentrations to such an extent that the process of glucose metabolism can be measured and analysed in a continuous manner and with sufficient time resolution. We found no commercial device which could provide such data.

The results of this pilot study are in accordance with the recent state of the art of development of closed loop systems using CGMS-augmented insulin pumps\textsuperscript{9,11,19,20}.

Example 1

a) Plasma glucose concentration is 3.5 mmol/L, i.e. it is close to hypoglycaemia.

b) The approximate highest trend of plasma glucose concentration decrease/increase is 0.2 mmol/L/min (as found in this study), i.e. 1 mmol/L in 5 min.

c) In the course of five minutes, glycaemia may either fall from 3.5 to 2.5 mmol/L (which is hypoglycaemia) or rise to 4.5 mmol/L (which is euglycaemia).

d) Which steps should be considered? It is important not to interpret the patient’s state as plasma glucose 3.5 mmol/L, but as the point of its trajectory with possible trends. It is improbable that 5 minutes delay would cause a substantial deterioration of the state of the patient and that the patient would approach severe hypoglycaemia. It is therefore appropriate to repeat the measurements after 5 min. The decision is made on the basis of the trend between the last two measurements. If plasma glucose concentration decreases and the trend decreases further it is necessary to immediately take glucose. If, however, the trend increases significantly, and after 5 min, the glucose concentration is 4 mmol/L, it is clear that the patient should remain at rest, and should not take up glucose. Administration of glucose in the case of low plasma glucose concentration and its increasing trend will definitely lead to hyperglycaemia because time to absorb
glucose varies from 10 to 15 min. The whole system starts to oscillate. In other words the compensation system has deteriorated.

Example 2
a) Plasma glucose concentration is 15 mmol/l, i.e. hyperglycaemia.

b) The approximate highest trend of plasma glucose concentration decrease/increase is 0.2 mmol/L/min (as found in this study), i.e. 1 mmol/L in 5 min.

c) In the course of five minutes, glycaemia may either fall to 14 mmol/L or rise to 16 mmol/L.

d) Which steps should be considered? Plasma glucose concentration should be measured after 5 min. If a decreasing trend is detected, it might be a mistake to add insulin to restore normoglycaemia. The addition of insulin might sometimes lead to overshoot to hypoglycaemia. For this reason, information on insulin available in the body, on food consumption and after muscular exercise is necessary, to assess the real needs for correction boluses. On the other hand, if, after 5 min, the glucose concentration increases above 15.5 - 16 mmol/L, the need for a correction bolus is evident.

The time has come to use CGM more widely in diabetes management and to introduce metrics that allow assessment of continuous glucose sensing for better glycem control on a day-to-day basis. Data analysis software as well as computer-assisted decision support systems have the potential to optimize clinical decision making. Challenging research is on going. Several sophisticated studies are described elsewhere:

The International Diabetes Center, Minneapolis, USA, has developed the data analysis software program called Ambulatory Glucose Profile (AGP) “Dashboard” and issued recommendations for standardizing glucose reporting and analysis to optimize clinical decision making. To date, 10-point glycaemic profiles are applied in our diabetes center to correct diabetes management.

To ease analysis of CGM and SMPG/SMBG data, Rodbard and Vigersky developed a computer-assisted decision support (CADS) for primary care providers to improve diabetes management in type 2 diabetes patients. This system is based on the input of SMBG/SMPG data, including clinical information.

Recent studies provide an optimistic outlook on perspectives for advanced technologies for the treatment of diabetes.

In summary, the practical outcomes from the repeated OGTTs in our 4-day pilot study in volunteers tested when lying in bed and keeping to the defined meal plan:

1) The time delay (lag phase) between glucose concentrations in plasma and tissue in the course of their increase/decrease was in HP 4-12 min/14-30 min, in DM1 0-18 min/27-37 min and in DM2 7-20 min/19-123 min. The delays appear to be longer in people with diabetes than in healthy persons. The decrease in glucose concentration always lasted longer than the increase.

2) The maximum glucose increase/decrease rate in HP < 0.1 mmol/L/min. In DM1 and DM2, the maximum glucose increase rate was 0.25 mmol/L/min/ and the maximum glucose decrease rate was mostly 0.15mmol/L/min with one extreme value of 0.25 mmol/L/min.

Hence, considering the assessment of various disorders of carbohydrate metabolism, the glucose increase/decrease rates appear to be more sensitive than oral GTT alone. Therefore, in addition to AGP, ketones, HbA1c and insulin sensitivity (HOMA), the metabolic state characteristics should be completed by information on lag-phase and maximum GID rates in GTT. Further studies considering the lag phase for the adjustment of euglycaemia-oriented therapy are underway.

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Authorship contributions: RC, JK, MOC: study design, data processing and interpretation, literature search, manuscript writing; BS, RP, LJ: preparation of study run, data collection and statistical analysis; TB: data processing, figures; BD: laboratory analysis, supervision of laboratory procedures and equipment; HZ, ED: meal plan and CGMS, blood data collection; JV: technical assistance, figures; JB: manuscript writing, literature search.
Conflict of interest statement: RC: participation in clinical studies on insulin pumps was supported by Medtronic Minimed International; MOC: employees, Probe Scientific Ltd, Bedford, United Kingdom; BS, JK, RP, LJ, JV: employee, BVT Technologies, a.s., Strazeck, Czech Republic; TB, BD, HZ, ED, JB: none conflict of interest.

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